

DNA-Bioprotective Effects of Lactic Acid Bacteria against Aflatoxin B₁

SEBNEM KURHAN^{1*} and IBRAHIM CAKIR²

¹Abant Izzet Baysal University, Novel Food Technologies Development, Application and Research Center, Bolu, Turkey.

²Abant Izzet Baysal University, Faculty of Engineering and Architecture, Department of Food Engineering, Bolu, Turkey.

<http://dx.doi.org/10.12944/CRNFSJ.4.Special-Issue-October.11>

(Received: August, 2016; Accepted: September, 2016)

ABSTRACT

Lactic acid bacteria commonly named as probiotics have a broad range of utilization area in human diet and food industry. Besides these known properties of probiotics, in recent years many researchers have focused on their anti-genotoxic called DNA-bioprotective effects. Human gets exposure with genotoxins such as mycotoxins, polycyclic aromatic hydrocarbons, and *n*-nitroso-compounds through diet and environmental contaminations. Aflatoxin B₁ is mycotoxin which was reported one of the most potent hepato-carcinogen and its exposure stems from human diet. In this study, we aimed to investigate DNA-bioprotective effect of *Lactobacillus plantarum* on human colon adenocarcinoma (Caco-2) cells against Aflatoxin B₁ (AFB₁) with comet assay without metabolic activation. The results showed that DNA-bioprotective effect of *L. plantarum* did reduce the AFB₁'s genotoxic effect on colon adenocarcinoma (Caco-2) cells. Positive control (50 μM H₂O₂ applied) and high dose (>10ppm) AFB₁ applied cells have the same comet tail appearance. Only visual scoring is performed. Besides negative control cells (Only PBS) and *Lactobacillus plantarum*+AFB₁ mixture showed the same manner with each other; no comet tail detected. These results clearly indicate that *L. plantarum* is capable of reduce AFB₁ safely without producing any by-products.

Keywords: Probiotics, aflatoxin B₁, *L. plantarum*, comet assay (SCGE).

INTRODUCTION

Lactic acid bacteria are a wide microorganism group which is known for formation of lactic acid as a dominant metabolite with their sugar metabolism (Zhong *et al.* 2014). Lactic acid fermentation is the most common and the oldest way of food preservation. Besides preservation aspects of lactic acid bacteria are seen from mouth to the end of the intestinal system of human and other mammals (Molin 2001, de Vries *et al.* 2006).

Lactic acid bacteria commonly named as probiotics have a broad range of utilization area in human diet and food industry. Probiotics are defined as "living microorganisms when consumed adequate amounts provide health benefit to the host" by FAO/WHO working group (FAO/WHO, 2002). The use of probiotics has an increasing demand for their prevention and treatment of diseases mainly intestinal diseases (Zhong *et al.* 2014). On the other hand, in recent years many researchers have focused on their anti-genotoxic called DNA-bioprotective effects.

Human gets exposure with genotoxins such as mycotoxins, polycyclic aromatic hydrocarbons, and *n*-nitroso-compounds through diet and environmental contaminations. Mycotoxins are the secondary metabolite of fungus. Production requirements change with ambient temperature, water activity, pH, utilizable nutrients and competition with the other micro-organisms (El-Nezami *et al.* 1998). Aflatoxin B₁ is mycotoxin which was reported one of the most potent hepato-carcinogen and its exposure stems from human diet. Once the food is contaminated with AFB₁, it can be removed physical and chemical treatments. However, these treatments may lead to undesirable properties of food such as loss of nutritional quality, flavor etc. and need expensive investments.

Recent years, there is a growing interest of decontaminating DNA-reacting compounds with microbial processes. The first reported study by Ciegler *et al.* (1966), indicate many kind of microorganisms, bacteria, yeasts, moulds, actinomycetes and algae are able to reduce aflatoxin in food and feed. DNA-bioprotective effect depends on probiotic strain (Cenci *et al.* 2005, Caldini *et al.* 2008). One strain may show the antigenotoxic activity efficiently, one strain may be the genotoxic itself. Mainly, antigenotoxic effect of these probiotic strains is a new functional property (Cenci *et al.* 2005).

In this study we aimed to investigate DNA-bioprotective effect of *Lactobacillus plantarum* on human colon adenocarcinoma (Caco-2) enterocytes against Aflatoxin B₁ (AFB₁) with comet assay without metabolic activation.

MATERIALS AND METHODS

Bacterial culture

Lactobacillus plantarum isolated from fermented food in previously study was maintained in deMan Rogosa Sharpe (MRS) Broth at 37°C for 18 hours in 5% CO₂ supplemented incubator. After 18 h incubation total viable lactic acid bacteria was determined with plate count method on MRS Agar incubated at 37°C for 24-48 h in CO₂ incubator.

Caco-2 cell culture

Human intestinal colon adenocarcinoma, Caco-2 cell line, was used in the study. Caco-2 cells

were grown in Dulbecco's Modified Eagle Medium F-12 (DMEM) supplemented with 10% (v/v) fetal bovine serum (FBS), 1% (v/v) L-glutamine, %1 (v/v) penicillin-streptomycin solution. The cells were maintained at 37°C in a 5% CO₂ atmosphere and sub-cultivated at 80% confluences with Trypsin-EDTA.

Cell viability test (Trypan blue dye exclusion test)

Cell viability was determined with trypan blue dye exclusion test which is based on the exclusion of the dye by the dead cells. Viable and non-viable cells were counted with Neubauer hemocytometer on the light microscope using x10 objectives.

Genotoxin preparation

Genotoxin, AFB₁, was prepared by following method by Topçu *et al.*(2010).

L. plantarum and AFB₁ co-incubation

L. plantarum was centrifuged 10000 rpm for 5 minutes and supernatant discarded. Pellet was washed twice with Ca⁺⁺ and Mg⁺⁺ free phosphate buffered saline (PBS). Pellet was solved in AFB₁ containing PBS and 5 mL of suspension placed in 6-well-plates and agitated 37°C for 6 hours on orbital microplate shaker. Negative control (PBS) and AFB₁ without *L. plantarum* placed the 6-well-plate either. *L. plantarum* viability and total number was assigned at both 0. and 6th hours.

Comet assay (Single cell gel electrophoresis)

Comet assay was performed as the Comet Assay datasheet manual with minor modifications (Enzo Life Sciences, New York, ABD). Cells were seeded on the 6-well-plate two days before the test. After 18 hours of seeding, cells were exposed to genotoxins for 16 hours. Test was conducted with positive and negative controls. For this purpose, 50 μM H₂O₂ was used as positive; PBS was used as negative control. Positive control was prepared just before the assay on the ice.

After genotoxin exposure, medium in the wells was discarded using sterile single-use Pasteur pipette, and washed with ice cold PBS to remove medium and metabolite residues and trypsinized the cells at 37°C for 5-7 minutes. Then cells were

dissolved in PBS and cell viability was checked. PBS+cell suspension was used for the Comet assay.

Cells were counted on the Neubauer hemocytometer; cell suspension was added in the low melting agarose (LMA), mixed well and transferred onto the comet slide. Slides were placed on a flat surface in the refrigerator. Agarose was hardened and slides were placed into the lysis solution. Slides were immersed cold lysis solution at least 2 hours. In the end of lysis, slides were washed with deionized water and put in the freshly prepared alkali solution. This step let cleave the double bonds in the alkali labile sites of DNA molecule. Slides were placed onto the electrophoresis tank and power supply was set 1 V/cm and current 300 mA for 40 minutes. Electrophoresis was conducted in alkali (pH>13) conditions. CyGreen nucleic acid dye use for imaging comets. x10000 stock dye solution diluted x10 concentration and slides were dyed. Comets

were screened using x10 objectives with FITC filter equipped Nikon C2 confocal laser scanning microscope.

RESULTS

L. plantarum-AFB₁ co-incubation

Lactic acid bacteria+genotoxin co-incubation did not reduce the viable lactic acid bacteria cell count.

Caco-2 cell viability

Comet test was started with same cell count (10⁵ cells). AFB₁ and negative control cells showed higher viability (77.5% and 81%, respectively) than AFB₁+LAB1 and positive control cells. This means current doses of AFB₁ did not lead any cytotoxic effect. However, when AFB₁ was together with *L. plantarum* cell viability was reduced (64%). This means lactic acid bacteria have cytotoxic effect on colon adenocarcinoma cells.

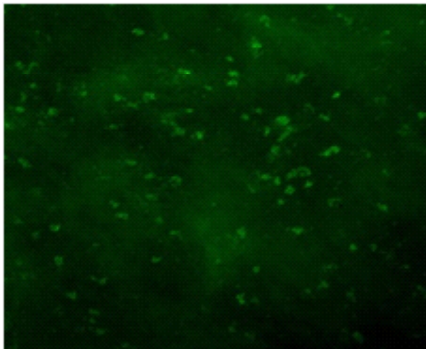


Fig. 1: Negative control- Caco-2 cells

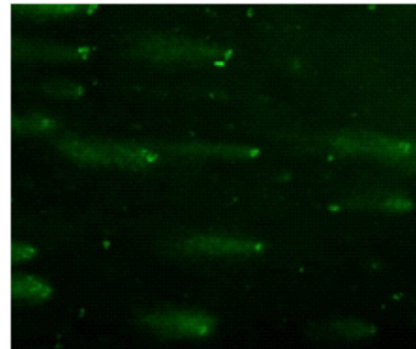


Fig. 2: Positive control- Caco-2 cells

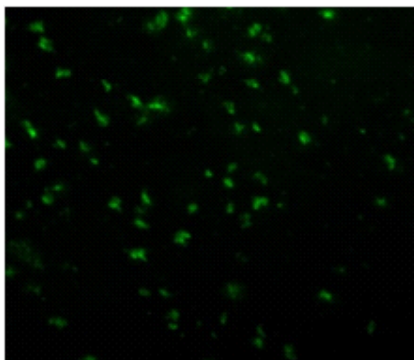


Fig. 3: LAB+AFB1 applied Caco-2 cells

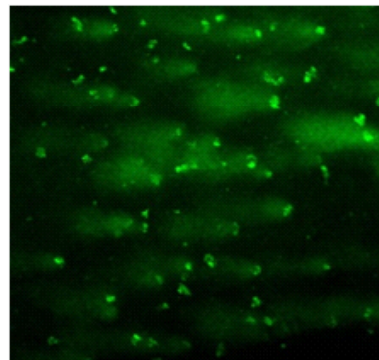


Fig. 4: Only AFB1 applied Caco-2 cells

L. plantarum+AFB₁ application to the Caco-2 cells caused to diminish Caco-2 cell size and viability (Data not shown). This result indicates cytotoxic effect of *L. plantarum* on Caco-2 cells. Similar results were taken by Er *et al.* (2015). They found that *L. plantarum*'s cell-free filtrate, after 24 hour incubation with Caco-2 cells, showed the highest (50% inhibition) cytotoxic effect.

Comet Assay

Comet assay results were scored visually. Images was categorized as; No-migration, low-migration, high-migration (Kadioglu *et al.* 2009). Samples were divided into 4 categories; negative and positive controls, AFB₁ and AFB₁+LAB applied groups. While no tail was observed in the negative controls and AFB₁+LAB group; positive and AFB₁ group was showed high migration in the tails.

Tail appearance in the alkaline comet assay indicates single stranded DNA breaks, double-stranded DNA breaks and majority of apurinic, apyrimidinic sites as well as alkali labile sites in the DNA molecule. Tail length is related with the damage quantity. *Lactobacillus plantarum* is able to reduce/bind AFB₁ 13-54% according to our HPLC study. Reducing amount by *Lactobacillus plantarum* is based on incubation period/time. The most efficient binding rate was found at 6th hour. LAB+AFB₁

complex is a reversible formation. After 6 hours, LAB+AFB₁ complex started to break down, and binding rate reduced. In spite of having uncertainty in the literature about by-products of AFB₁ that was eliminated with lactic acid bacteria, our results showed that *Lactobacillus plantarum* is able to reduce to AFB₁ without any toxic by-products.

DISCUSSION

While *Lactobacillus rhamnosus* and *L. paracasei* are abundant in dairy products, *L. plantarum* generally is responsible for plant originated fermentation. *L. plantarum* play a key role especially olive and sauerkraut fermentations (de Vries *et al.* 2006). Thus, human consume fermented plant products also consume substantial amount of *L. plantarum* (Molin 2001). To enhancement of diet with fermented food is easy and healthy way for protecting genotoxins.

ACKNOWLEDGEMENTS

This project was funded by Abant Izzet Baysal University, Scientific Research Projects Coordination Unit (Project number: 2015.09.04.843). We would like to thank Abant Izzet Baysal University, Novel Food Technologies Development, Application and Research Center (YENIGIDAM)

REFERENCES

1. Caldini, G., Trotta, F., Villarini, M, Moretti, M., Pasquini, R., Scassellati Sforzolini, G. Cenci, G. Screening of Potential Lactobacilli Antigenotoxicity by Microbial and Mammalian Cell-Based Tests. *International Journal of Food Microbiology*, **102**: 37-47: 2005.
2. Caldini, G., Trotta, F., Corsetti, A., Cenci, G. Evidence for In Vitro Anti-genotoxicity of Cheese Non-starter Lactobacilli. *Antonie van Leeuwenhoek*, **93**: 51-59: 2008
3. Cenci, G., Caldini, G. and Trotta, F. Inhibition of DNA Reactive Agents by Probiotic Bacteria. *Recent Research Developments in Applied Microbiology & Biotechnology*, **2**: 103-121: 2005.
4. Ciegler, A., Lillehoj, E.B., Peterson, R.E., Hall, H.H. Microbial Detoxification of Aflatoxin. *Applied Microbiology*, **14**(6): 934-939: 1966.
5. De Vries, M.C., Vaughan, E.E., Kleerebezem, M., de Vos, W.M. *Lactobacillus plantarum*-Survival, Functional and Potential Probiotic Properties in the Human Intestinal Tract. *International Dairy Journal*, **16**: 1018-1028: 2006.
6. El-Nezami, H, Kankaanpaa, P, Salminen, S, Ahokas, J. Ability of Dairy Strains of Lactic Acid Bacteria to Bind a Common Food Carcinogen, Aflatoxin B1. *Food and Chemical Toxicology*, **36**: 321-326: 1998.
7. Er, S., Koparal, A.T. and K yvan , M. Cytotoxic Effects of Various Lactic Acid Bacteria on Caco-2 Cells. *Turkish Journal of Biology*, **39**:

- 23-30: 2015.
8. Fuchs, S., Sontag, G., Stidl, R., Ehrlich, V., Kundi, M., Knasmüller, S. Detoxification of Patulin and Ochratoxin A, Two Abundant Mycotoxins, by Lactic Acid Bacteria. *Food and Chemical Toxicology*; **46**: 1398-1407: 2008.
 9. Guidelines for the evaluation of probiotics in food, London Ontario, Canada. April 30 and May 1, 2002.
 10. Kadyoğlu, E., Sardas, S., Ertürk, S., Ozatamer, O., Karakaya, A.E. Determination of DNA Damage by Alkaline Halo and Comet Assay in Patients Under Sevoflurane Anesthesia. *Toxicology and Industrial Health*; **25**: 205-212: 2009.
 11. Molin, G. Probiotics in Foods not Containing Milk or Milk Constituents, with Special Reference to *Lactobacillus plantarum*. *The American Journal of Clinical Nutrition* ; **73** (suppl):380S–5S, 2001.
 12. Peltonen, K., El-Nezami, H., Haskard, C., Ahokas, J., Salminen, S. AflatoxinB1 Binding by Dairy Strains of Lactic Acid Bacteria and Bifidobacteria. *Journal of Dairy Science.*; **84**: 1256–2152: 2001.
 13. Topçu, A., Bulat, T., Wishah, R., Boyacı, İH. Detoxification of Aflatoxin B1 and Patulin by *Enterococcus faecium* Strains. *International Journal of Food Microbiology*; **139**: 202-205: 2010.
 14. Zhong, L., Zhang, X., and Covasa, M. Emerging Roles of Lactic Acid Bacteria in Protection Against Colorectal Cancer. *World Journal of Gastroenterology*; **20** (204): 7878-7886: 2014.